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Arsenic Exposure and Prevalence of the Varicella Zoster Virus in the United States: NHANES (2003-2004 and 2009-2010)

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Abstract

Background: Arsenic is an immunotoxicant. Clinical reports observe the reactivation of varicella zoster virus (VZV) in people who have recovered from arsenic poisoning and in patients with acute promyelocytic leukemia that have been treated with arsenic trioxide.

Objective: To evaluate the association between arsenic and the seroprevalence of VZV immunoglobulin G (IgG) antibody in a representative sample of the US population.

Methods: We analyzed data from of 3,348 participants of the National Health and Nutrition Examination Survey (NHANES) 2003-2004 and 2009-2010 pooled survey cycles. Participants were eligible if they were 6-49 years of age with information on both VZV IgG and urinary arsenic concentrations. Total urinary arsenic (TUA1) was defined as the sum of arsenite, arsenate, monomethylarsonic acid and dimethylarsinic acid. TUA2 was defined as total urinary arsenic minus arsenobetaine and arsenocholine.

Results: The overall weighted seronegative prevalence of VZV was 2.2% for the pooled NHANES sample. The geometric means of TUA1 and TUA2 were 6.57 $\mu\text{g/L}$ and 5.64 $\mu\text{g/L}$. After adjusting for age, gender, race, income, creatinine, and survey cycle, odds ratios for a negative VZV IgG result in association with 1-unit increases in $\ln\text{-TUA1}$ and $\ln\text{-TUA2}$ were 1.87 (95% CI: 1.03, 3.44) and 1.40 (95% CI: 1.0, 1.97), respectively.

Conclusions: In this cross-sectional analysis urinary arsenic was inversely associated with VZV IgG seroprevalence in the US population. This finding is in accordance with clinical observations of zoster virus reactivation from high doses of arsenic. Additional studies are needed to confirm the association and evaluate causal mechanisms.

Introduction

Initial exposure to the varicella zoster virus (VZV) causes chickenpox (varicella). Upon recovery from the initial infection, VZV establishes latency and if reactivated, it will cause shingles (herpes zoster) (Gershon et al. 2010). As long as the host maintains sufficient VZV-specific cell mediated immunity, the virus can remain latent indefinitely. Approximately 1 out of 3 people in the U.S. will develop herpes zoster (HZ) in their lifetime resulting in over one million cases every year (CDC 2011a; Gershon et al. 2010;). The risk of HZ increases in the elderly and in people with immunosuppressive illnesses and/or taking immunosuppressive medications (CDC 2011a). Other risk factors for HZ include diabetes, female gender, caucasian race, medical trauma and psychological stress (Gershon et al. 2010).

Interestingly, medical case reports dating back to the 1900s have documented the appearance of fine punctate irritating rashes, herpetic skin-eruptions, and HZ in people shortly after they recovered from acute and sub-chronic arsenic poisoning (Bartolome et al. 1999; Carlill 1917; Hope-Simpson 1965; Jacob 1931; Satterlee 1960; Uede and Furukawa 2003 Walsh 1900). More recently, clinicians have identified HZ as a common side effect for patients with acute promyelocytic leukemia (APL) after they received treatment with arsenic trioxide (Au and Kwong 2005; Douer and Tallman 2005; Nouri et al. 2006; Rousselot et al. 2004; Subbarayan et al. 2007; Tanvetyanon and Nand 2004) While no study has been conducted to determine if HZ is more common among APL patients who receive arsenic trioxide versus other treatments, physicians commented that only the patients treated with arsenic trioxide developed HZ (Tanvetyanon and Nand 2004). There is evidence from animal models and *in vitro* studies that arsenic exposure can alter the immune response (Dangleben et al. 2013). However, it is unknown if exposure to arsenic from environmental sources affects the VZV immune response.

Currently, two National Health and Nutrition Examination Surveys (NHANES) included both VZV IgG serology testing and urinary arsenic measurements. Using these data, we sought to determine if arsenic exposure in the general U.S. population is a risk factor for VZV IgG seronegativity.. Our hypothesis was that higher urinary arsenic concentrations would be associated with a higher seronegative prevalence of VZV IgG.

Methods

Study population

The National Health and Nutrition Examination Survey (NHANES) is designed to be a nationally representative sample of the resident, non-institutionalized US civilian population. Participation rates for the 2003-2004 cycle and the 2009-2010 cycles were 76% and 77.3% respectively (CDC 2013a). Informed consent was obtained from all survey participants and all study protocols were approved by the National Center for Health Statistics research ethics review board (CDC 2013b).

In this study, we restricted our analyses to individuals who had data on both VZV serology and urinary arsenic speciation. Serological testing for VZV antibodies was performed on participants aged 6-49 years. Urinary arsenic concentrations were measured in a random sample of one-third of the participants aged ≥ 6 years (Caldwell et al. 2009). This resulted in data on 1,641 individuals from the 2003-2004 survey and 1,718 individuals from the 2009-2010 survey. We excluded individuals with a positive test for human immunodeficiency virus (HIV) (n=4 for 2003-2004 and n=7 for 2009-2010) due to their compromised immune status. This resulted in a final sample size of 3,348 for the two pooled survey cycles.

Varicella Zoster Virus (VZV) serology

Initially, NHANES utilized an enzyme immunoassay method which detects disease-acquired immunoglobulin G (IgG) antibodies to VZV but is less sensitive to the IgG response produced by the vaccine (Reynolds et al. 2010). Subsequently, NHANES adopted a glycoprotein-based enzyme-linked immunosorbent assay that is more sensitive and reliably detects both vaccine and disease induced immunity (Hammond et al. 2006). We utilized all available VZV data to ascertain seroprevalence status across survey cycles to identify 98 seronegative and 3250 seropositive individuals that were used in the combined analysis.

NHANES 2003-2004

Varicella Zoster Virus IgG was measured in serum using a whole cell enzyme immunoassay (EIA). This assay was developed by the Immunoserology Unit of the California State Department of Health Services (CSDHS) Viral and Rickettsial Disease Laboratory (VRDL) and has been previously described (Forghani et al. 1978). This protocol yields an index of optical density (O.D.) reading where an $O.D. \geq 1$ indicates the presence of the VZV antibody (seropositive) and an $O.D. < 1$ indicates that the antibody was not detected (seronegative). A negative IgG VZV test is useful to determine whether an individual is susceptible to infection by the virus (CDC 2011b). Subsequently the CDC re-tested a subsample on this survey cycle using a glycoprotein-based enzyme-linked immunosorbent assay (gp-ELISA), which is considered to have higher sensitivity and specificity compared to the EIA method, and determined that the EIA assay produced false-negative results for 26% of participants aged 6-19 years (Reynolds et al. 2010). NHANES made the gp-ELISA data available in 2009. Subsequently, we used data from the both the EIA assay and the available gp-ELISA assay to classify VZV status in this cycle. Specifically, the EIA and the gp-ELISA methods agreed on 40 negative and 37 positive VZV

individuals that were re-tested. Sixteen individuals were re-classified as seropositive based on the gp-ELISA results that were initially identified as VZV negative by the EIA assay. Finally, 14 individuals with an equivocal gp-ELISA result were left as initially identified by the EIA method. This resulted in 69 seronegative and 1,568 seropositive individuals for this cycle.

NHANES 2009-2010

In this survey the CDC evaluated the presence of the VZV antibody using an enzyme immunoassay (EIA) as previously described. However, all samples that tested negative (O.D. <1) or in the equivocal range were re-evaluated using the gp-ELISA method. Therefore, all negative IgG VZV results in this survey were confirmed, yielding greater accuracy for the serological classification. This resulted in 29 seronegative and 1,682 seropositive individuals in this cycle.

Urinary arsenic assessment

Urinary arsenic concentrations are measured in a spot sample collected during the physical examination and analyzed within 3 weeks of collection using high performance liquid chromatography (HPLC) coupled to an inductively coupled-plasma dynamic reaction cell-mass spectrometry (ICP-DRC-MS). Seven urinary arsenic species were evaluated with this method: arsenite (As3), arsenate (As5), arsenobetaine (AsB), arsenocholine (AsC), monomethylarsonic acid (MMA), dimethylarsenic acid (DMA), and trimethylarsine oxide, as well as total urinary arsenic (CDC 2007). The corresponding limits of detection (LOD) for the urinary arsenic species used were 0.4 µg/L, 0.6 µg/L, 1.2 µg/L, 1.0 µg/L, 0.9 µg/L and 1.7 µg/L for AsB, AsC, As3, As5, MMA and DMA respectively. The LOD for total urinary arsenic in the 2003-2004 was 0.6 µg/L and changed in the 2009-2010 cycle to 0.74 µg/L. Samples with arsenic measurements below the LOD were assigned a level equal to the LOD divided by the square root of two. The proportion of samples below the LOD in 2003-2004 was 44.57% for AsB, 98.9% for AsC,

93.28% for As3, 92.61% for As5, 60.26% for MMA, 11.46% for DMA, and 1.49% for total arsenic. In 2009-2010, the proportion of samples below the LOD was 37.14% for AsB, 98.2% for AsC, 94.97% for As3, 97.29% for As5, 67.39% for MMA, 19.93% for DMA, and 0.77% for total arsenic.

Arsenobetaine and arsenocholine are arsenosugars present in seafood and are considered to be non-toxic (Murer et al. 1992). Subsequently, we defined total urinary arsenic (TUA) using two approaches. The first approach (TUA1) was defined as the sum of As3, As5, MMA and DMA. The second approach (TUA2) was defined as total arsenic minus AsB and AsC. Since NHANES includes multiple demographic groups, it is recommended that urinary analyte concentrations should be evaluated as an unadjusted analyte concentration and have urinary creatinine as a separate independent variable to account for differences in urine dilution (Barr et al. 2005).

Adjustment variables

Variables that were considered in the analyses *a priori* as potential confounders for the association between VZV status and urinary arsenic concentrations included: age, sex, race/ethnicity, family poverty-income ratio, body mass index (BMI), urinary creatinine levels and survey cycle. Race/ethnicity was self-reported as non-Hispanic white, non-Hispanic Black, other Hispanics, Mexican American and other race including multiracial. Other Hispanics were collapsed into a single category with the other race category including multiracial for both survey cycles. BMI was calculated by dividing measured weight in kilograms by measured height in meters squared. BMI was classified as underweight (below 18.5), normal (18.5-24.9), overweight (25-29.9) and obese (30 and above). For participants that were <20 years of age BMI classification was defined using the CDC growth charts for age and sex specific cutoffs. Urinary creatinine was right skewed and subsequently natural log-transformed.

Statistical analysis

All statistical analyses were performed in Stata using the survey command to account for the complex sampling design (Version 12.1, StataCorp LP, College Station, Texas). Unweighted sample sizes are presented along with weighted prevalence and geometric means (GM) for each covariates. Standard errors (SEs) and confidence intervals (CIs) were estimated using the Taylor-linearization method. The statistical significance level was set at $\alpha=0.05$ and all statistical tests were two-tailed. The association was evaluated for the two survey cycles, independently. The two survey cycles were also combined to increase the precision for the estimated relationship and survey weights were re-scaled to match the US population at midpoint for the combined survey cycles.

Total urinary arsenic concentrations (TUA1 and TUA2) were right skewed and natural log-transformed for the analyses. Linear regression models were used to calculate the GM and SE of TUA by prevalence of VZV status and all covariates evaluated, for both survey cycles and for the combined sample. Associations between all covariates and urinary arsenic were evaluated using a Wald test for significance to evaluate the overall association. The models estimating the GM of TUA for each strata always included urinary creatinine.

Logistic regression models were used to evaluate the association between seronegative VZV status and \ln TUA, for the combined sample and the two survey cycles independently. This approach included all *a priori* covariates that were considered risk factors for VZV (e.g. age, sex, race/ethnicity, family poverty income ratio, BMI), NHANES survey year, and \ln urinary creatinine. For the combined sample we tested the interaction between survey cycle and arsenic exposure in the adjusted model. These interactions were non-significant for TUA1 ($P=0.68$) and TUA2 ($P=0.39$), which suggested that the associations did not differ by survey year and further

supported combining the two survey cycles. As a sensitivity analysis we also analyzed these logistic regression models without the sample weights (unweighted). This unweighted approach would prevent spurious associations that could result if a heavy weight was attached to few individuals. Further, weighted penalized splines were used to evaluate non-linear relationships with knots at the 5th, 25th, 50th and 95th percentile of exposure. Since weighted penalized splines using the pooled data may produce unreliable 95% confidence intervals, we used generalized additive models to evaluate non-linear relationships and compute 95% confidence intervals without including survey weights.

Results

A total of 3,348 participants had arsenic measurements defined as TUA1 and a total of 3,283 participants had arsenic measurements defined as TUA2 across both survey cycles. The difference in sample size (n=65) was due to samples missing total urinary arsenic measurements used in compute TUA2 as compared to the metabolites used compute TUA1. There were 98 individuals who were seronegative for the Varicella-Zoster Virus (VZV) IgG in the pooled study sample. The overall geometric mean of total urinary arsenic for the pooled sample was 6.57 µg/L (95% CI: 6.26, 7.91 µg/L) and 5.64 µg/L (95% CI: 5.20 to 6.12) for TUA1 and TUA2, respectively. The population characteristics for the pooled sample are provided in Table 1 and differ by 65 participants based on the exposure assessment approach used to estimate total urinary arsenic as a result of missing total urinary arsenic measurements. Total urinary arsenic was significantly associated with race/ethnicity, age, and VZV serology based the overall creatinine adjusted Wald test (Table 2). In the pooled sample, TUA1 and TUA2 were slightly higher among VZV seronegative IgG participants compared to seropositive individuals (8.31 µg/L versus 6.77 µg/L, p=0.01; and 7.62 µg/L versus 5.85 µg/L, p=0.02). Only TUA1 was

associated with BMI classification. The geometric means of total urinary arsenic, which were adjusted for creatinine, was not different by sex or family poverty-income ratio (Table 2).

In adjusted models, odds ratios for a negative VZV IgG result in association with 1-unit increases in \ln -TUA1 and \ln -TUA2 were 1.87 (95% CI: 1.03, 3.44) and 1.40 (95% CI: 1.0, 1.97), respectively for the pooled survey sample (Table 3). These models were adjusted for age, \ln urinary creatinine, sex, race/ethnicity, family poverty-income ratio, BMI classification and survey cycle. Since this is a relatively small sample size and there was missing data, we also analyzed the association between arsenic and VZV-status using a more parsimonious approach that only controlled for age, survey cycle and urinary creatinine. These models with less missing data yielded consistent results where the odds of having a negative VZV result increased with each increase in \ln TUA1 (aOR: 2.24, 95% CI: 1.37, 3.56) and TUA2 (aOR: 1.58, 95% CI: 1.17-2.13). Furthermore, the unweighted adjusted models yielded consistent results suggesting that the observed association was not driven by spurious data that can result if a few individuals are given a greater weight. Finally, additional analysis that examined the association between total urinary arsenic and VZV seroprevalence in each survey cycle independently yielded a consistent association between negative VZV IgG seroprevalence and arsenic (Table 3).

The shape of the dose-response relationship between total urinary arsenic and VZV seronegative status was modeled using penalized splines as both weighted and unweighted samples (Figure 1). The shape of the dose-response relationships was approximately linear for unweighted spline models of TUA1 and TUA2, Figure 1A and 1B respectively. The weighted dose-response curves were in comparison more attenuated and non linear for TUA1 and TUA2, Figure 1C and 1D respectively. Despite these subtle differences in the shape of the dose-response curve between

arsenic and VZV seronegative status, the weighted and unweighted analyses yielded a consistent positive association.

Discussion

In a representative sample of the U.S. population aged 6 to 49 years, higher concentrations of total urinary arsenic were associated with a higher prevalence of negative VZV serology results after adjusting for other risk factors. While the shape of the dose-response relationship could have been biased by accounting for urinary metabolites that were below the limit of detection, a significant positive association was observed in two NHANES surveys taken six years apart where the odds of a negative VZV result increased approximately 40-95% for each unit increase in natural log-transformed urinary arsenic concentrations (TUA1 or TUA2). The association was consistent in the pooled analytic sample and across each survey cycle. These findings, which are the first to look at the association between arsenic exposures and VZV status in the United States general population and across a relatively modest range of exposures, builds upon prior experimental evidence and clinical observations that shows that therapeutic doses of inorganic arsenic affect and/or suppress specific immune functions and support the hypothesis that arsenic exposure diminishes VZV immunity.

Arsenic is a common environmental contaminant that can be found in groundwater and in the food chain. In this study, it was not possible to determine the route of arsenic exposure or the species of arsenic that comprises the exposure because only a spot urine sample was collected for arsenic measurement. However, in the U.S., arsenic contaminated drinking water is considered to be the dominant source of exposure and is mostly a concern for communities that rely on groundwater as their source of potable water (US EPA 2001). A 2001 study that used data collected by the EPA, estimated that 34 million Americans were drinking water with average

arsenic concentrations above 50 µg/L which was the maximum contaminant level (MCL) for drinking water at that time (Mushak 2000). In 2001, the EPA lowered the MCL to 10 µg As/L due to concerns about elevated risk of internal cancers and municipalities had until 2006 to comply (Abedin et al. 2002). Private drinking water wells, however, are not monitored or regulated by the EPA and survey data suggests that 11% - 19% of private wells exceed 10 µg As/L (Focazio et al. 2006; Montgomery et al. 2003; Twarakavi and Kaluarachchi 2006). Crops, particularly rice and cereal grains, can also take up arsenic from the soil and irrigation water (Cascio et al. 2011). Although dietary sources of arsenic are receiving more attention as a result of data from recent studies showing that people who regularly consume rice have higher urinary arsenic levels (Davis et al. 2012). Subsequently, it is possible that the lower exposure levels measured in this study resulted from dietary intake. Also, while we were able to calculate a total urinary arsenic concentration that did not include non-toxic arsenosugars, the small sample size prohibited us from examining the association between VZV-status and individual urinary arsenic metabolites.

There have been many clinical reports of VZV re-activation among people who have survived acute arsenic poisoning and among patients with acute promyelocytic leukemia that have been treated with arsenic trioxide (Au and Kwong 2005; Bartolome et al. 1999; Douer and Tallman 2005; Hope-Simpson 1965; Rousselot et al. 2004; Tanvetyanon and Nand 2004; Nouri et al. 2006; Subbarayan et al. 2007). Yet the biological mechanisms by which this occurs are not well understood. Researchers have shown that arsenic trioxide prevents the loss of virions from the perinuclear cell region, leading to an increase in the cellular vector genome retention (Mitchell et al. 2013). There is also considerable evidence from *in vitro* and *in vivo* experimental studies that inorganic arsenicals are potent immunotoxicants. High doses of inorganic arsenic are known to

suppress IgM and IgG antibody formation, inhibit antigen-driven T-cell proliferation and macrophage activity, block the differentiation of monocytes into functional macrophages, decrease CD4⁺ splenic cell numbers, and alter the development, activation and proliferation of T-cells (Bourdonnay et al. 2009; Burchiel et al. 2009; Burns and Munson 1993; Conde et al. 2007; Dangleben et al. 2013; De La Fuente et al. 2002; Galicia et al. 2003; Hernandez-Castro et al. 2009; Kozul et al. 2009a; Lemarie et al. 2006a; Lemarie et al. 2006b; Martin-Chouly et al. 2011; Nain and Smits 2012; Patterson et al. 2004; Sikorski et al. 1989; Soto-Pena et al. 2006; Yoshida et al. 1987;). Additionally, data from experimental models shows that high doses of inorganic arsenic influence viral pathology (Dangleben et al. 2013; Kozul et al. 2009b; Martin-Chouly et al. 2011; Mitchell et al. 2013; Patterson et al. 2004; Ramsey et al. 2013; Sebastian et al. 2006). Less is known about the effects of methylated arsenical species on immunological outcomes.

The best way to reduce the risk of chicken pox and HZ is to be vaccinated for VZV. Since 1996, the Advisory Committee on Immunization Practices has recommended routine VZV vaccination of all children aged 18 months to 2 years, susceptible adolescents, and adults that are at high risk of exposure to the virus (Marin et al. 2007). The vaccine contains live attenuated VZV which is very effective and induces immunity in >95% of the people who receive the vaccine (Reynolds et al. 2010). The vaccine results in latent infection which can be reactivated and cause HZ although studies have shown that the risk of reactivation after the vaccine is lower than for people who were infected with wild-type VZV (Reynolds et al. 2010). It would be useful if future studies considered whether the As-seroprevalence VZV association is only in those with wild-type VZV as compared to people who have been vaccinated against VZV.

Important strengths for this study include the use of a representative sample of the U.S. population exposed to arsenic at environmental concentrations. We also used urinary biomarkers to assign personal exposure levels of arsenic. Further we adjusted for relevant risk factors for VZV and urinary arsenic concentrations. The rigorous quality control procedures implemented in NHANES is also an important strength of the quality of the data presented. Although the protocol for the VZV assay changed between cycles, there was improved precision in the data collected in 2009-2010 which is likely a function of the glycoprotein-based enzyme-linked immunosorbent assay for determining VZV status. This assay has a higher sensitivity and specificity for VZV antibodies produced by vaccination which likely accounts for the more precise odds ratios in this cycle even though the actual number of VZV-seronegative samples was lower. Finally, the reproducibility of the effect and the observation of an exposure-response relationship between increasing total urinary arsenic concentration and VZV serology in two different survey cycles is reassuring.

There are also limitations to this study that must be considered. NHANES is a cross-sectional study and the temporality between arsenic exposure and VZV serology cannot be assessed. NHANES only measures urinary arsenic concentrations in one-third of the participants and urinary arsenic metabolites have a relatively short half –life which limits their use for estimating historical or long term exposures (Buchet et al. 1981). The presence of unmeasured confounders cannot be ruled out since serum specific IgG response resulting from vaccination or natural infection could be modified by other immune suppressive conditions or chronic infections. Yet we accounted for several important risk factors in the population including age, race, gender and BMI. Also, the seronegative VZV prevalence is low in both cycles for our combined sample. Misclassification of participants that were infected with VZV previously but did not mount a

sufficient IgG response that could allow for accurate serology testing is also possible. Furthermore, a negative VZV IgG result could also indicate that an individual has not been previously infected with the virus and that arsenic exposure was protective. This interpretation, however, seems unlikely since arsenic is known to be immunotoxic, the clinical reports of VZV re-activation after arsenic exposure, estimates that 99.5% of people born in the U.S. who are ≥ 40 years have had VZV, and the widespread adoption of VZV vaccines after 1996 (CDC 2012). Unfortunately, there was no available information on other antibodies such as IgM to improve our classification of VZV status. We were also not able to evaluate the observed association in children younger than 6 years of age or adults older than 45 years of age. VZV reactivation is a common problem among older adults with the incidence of zoster increasing at age 50-60 years, so the impact of arsenic exposure on this group may be different (Harpaz et al. 2008).

Conclusions

Total urinary arsenic concentration was positively associated with seronegative VZV IgG prevalence in a population with modest arsenic exposure. This information builds upon experimental studies and clinical observations showing an association between acute exposures to high levels of arsenic with herpes zoster and supports a link between environmentally-relevant levels of arsenic exposure and VZV- specific immune response. Additional studies are needed to more fully evaluate the effect of arsenic on other parameters of immune functioning and its ability to cause VZV reactivation. From a public health perspective, confirmation of arsenic's ability to suppress specific immune functioning has important implications for vaccine-preventable illnesses.

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Table 1. Population characteristics for the combined NHANES 2003-2004 and 2009-2010. Un-weighted sample sizes and weighted percentages.

Characteristic	TUA1^a	TUA2^b
Total Sample Size (N)	3,348 n (%)	3,283 n (%)
Sex		
Male	1,673 (50.3%)	1,649 (50.6%)
Female	1,675 (49.7%)	1,634 (49.4%)
Race		
Non-Hispanic white	1,215(62.7%)	1,177 (62.3%)
Non-Hispanic black	845 (12.9%)	839 (13.1%)
Mexican Americans	868 (12.1%)	853 (12.2%)
Other/Other Hispanics	420 (12.3%)	414 (12.4%)
Family Poverty Income Ratio		
Below poverty level (≤1)	943 (19.6%)	926 (80.2%)
Above poverty level (>1)	2,185(81.4%)	2,139 (19.8%)
Missing	220	218
BMI (kg/m²)		
Underweight	86 (2.2%)	85 (2.2%)
Normal	1,498 (41.2%)	1,469 (41.0%)
Overweight	825 (28.0%)	813 (28.1%)
Obese	909 (28.6%)	916 (28.7%)
Missing	30	23
Age (years)		
6-11	514 (11.5%)	534 (11.1%)
12-19	1,002 (17.6%)	1,063 (17.8%)
>20	1,598 (70.9%)	1,686 (71.1%)
TUA1^a (µg/L)		
Tertile 1(<4.8)	1,032 (33.7%)	----
Tertile 2 (>4.8-7.5)	1,116 (32.3%)	----
Tertile 3 (>7.5-139)	1,200 (34.0%)	----
TUA2^b (µg/L)		
Tertile 1(≤3.7)	----	1,004 (33.0%)
Tertile 2 (>3.7-8.4)	----	1,132 (33.2%)
Tertile 3 (>8.4-300)	----	1,147 (33.8%)
Creatinine (mg/dL)		
Tertile 1 (<80)	960 (31.6%)	917 (30.4%)
Tertile 2 (>80-153)	1,122 (32.6%)	1,113 (33.1%)
Tertile 3 (>153-768)	1,263 (35.8%)	1,253 (36.5%)
Varicella virus IgG		
Seropositive	3,250 (97.8%)	3,187 (97.8%)
Seronegative	98 (2.2%)	96 (2.9%)

^aTUA1= As3 + As5 + MMA + DMA. ^bTUA2 = Total As - Asc - AsB

Table 2. Weighted geometric mean (GM) and standard error (SE) of total urinary arsenic ($\mu\text{g/L}$) by demographic characteristics in the NHANES 2003-2004 cycle, 2009-2010 cycle, and a pooled sample (2003-2004 and 2009-2010).

NHANES Cycle	2003-2004	2003-2004	2003-2004	2003-2004	2009-2010	2009-2010	2009-2010	2009-2010	Pooled	Pooled	Pooled	Pooled
Exposure	TUA1 ^a	TUA1 ^a	TUA2 ^b	TUA2 ^b	TUA1 ^a	TUA1 ^a	TUA2 ^b	TUA2 ^b	TUA1 ^a	TUA1 ^a	TUA2 ^b	TUA2 ^b
	GM (SE)	P-value ^c	GM (SE)	P-value ^c	GM (SE)	P-value ^c	GM (SE)	P-value ^c	GM (SE)	P-value ^c	GM (SE)	P-value ^c
Seronegative (N)	69		69		29		29		98		98	
Characteristic												
Sex		0.27		0.94		0.09		0.01		0.58		0.15
Male	6.91 (1.05)		5.80 (1.09)		6.30 (1.03)		5.33 (1.04)		6.67 (1.03)		5.64 (1.05)	
Female	6.96 (1.03)		5.83 (1.08)		6.68 (1.04)		5.94 (1.05)		6.77 (1.03)		5.95 (1.05)	
Race		<0.001		<0.001		<0.001		<0.001		<0.001		<0.001
Non-Hispanic white	6.42 (1.04)		5.23 (1.09)		5.92 (1.03)		5.28 (1.04)		6.23 (1.03)		5.22 (1.06)	
Non-Hispanic black	6.52 (1.04)		5.63 (1.07)		6.00 (1.04)		5.22 (1.07)		6.31 (1.03)		5.40 (1.05)	
Mexican Americans	7.89 (1.05)		6.85 (1.05)		7.19 (1.03)		5.77 (1.05)		7.34 (1.03)		6.25 (1.03)	
Other/Other Hispanics	10.79 (1.06)		11.76 (1.10)		10.66 (1.10)		10.42 (1.08)		10.22 (1.05)		11.06 (1.07)	
Family Poverty Income Ratio		0.69		0.95		0.54		0.15		0.53		0.39
Below poverty level (<1)	6.87 (1.03)		5.84 (1.08)		6.40 (1.03)		5.84 (1.04)		6.69 (1.02)		5.81 (1.05)	
Above poverty level (>1)	7.00 (1.07)		5.87 (1.10)		6.55 (1.04)		5.34 (1.06)		6.82 (1.04)		5.60 (1.06)	
BMI (kg/m²)		0.002		0.36		0.70		0.05		0.03		0.19
Underweight	7.16 (1.09)		6.11 (1.08)		6.41 (1.16)		6.44 (1.15)		6.99 (1.06)		5.96 (1.04)	
Normal	7.17 (1.04)		6.11 (1.08)		6.52 (1.03)		5.82 (1.06)		6.85 (1.03)		6.97 (1.04)	
Overweight	6.93 (1.05)		5.84 (1.09)		6.56 (1.04)		6.20 (1.04)		6.86 (1.04)		5.94 (1.05)	
Obese	6.46 (1.03)		5.51 (1.07)		6.33 (1.03)		5.50 (1.04)		6.43 (1.02)		5.50 (1.04)	
Age (years)		0.002		<0.001		<0.001		<0.001		<0.001		<0.001
6-11	7.51 (1.06)		6.78 (1.11)		7.25 (1.03)		5.87 (1.06)		7.27 (1.04)		6.26 (1.06)	
12-19	6.41 (1.05)		4.66 (1.07)		5.60 (1.03)		3.96 (1.05)		5.90 (1.03)		4.29 (1.04)	
≥20	7.27 (1.04)		5.93 (1.08)		6.62 (1.03)		6.15 (1.04)		6.82 (1.03)		6.03 (1.04)	
Varicella virus IgG^a		0.08		0.06		0.004		0.02		0.01		0.02
Seropositive	7.09 (1.04)		6.16 (1.07)		6.47 (1.03)		5.61 (1.04)		6.77 (1.02)		5.85 (1.04)	
Seronegative	8.61 (1.12)		8.21 (1.17)		8.18 (1.07)		7.62 (1.13)		8.31 (1.08)		7.62 (1.12)	

^aTUA1= As3 + As5 + MMA + DMA. ^bTUA2 = Total As - AsC – AsB. ^cModels adjusted for ln transformed creatinine.

Table 3. Adjusted odds ratios (95% CI) for seronegative VZV status in association with 1-unit increase in ln-transformed total urinary arsenic (µg/L) for the combined NHANES sample (2003-2004 and 2009-2010).

NHANES Cycle	2003-2004 Weighted^a Odds Ratio (95% CI)	2003-2004 Unweighted^a Odds Ratio (95% CI)	2009-2010 Weighted^a Odds Ratio (95% CI)	2009-2010 Unweighted^a Odds Ratio (95% CI)	Pooled Weighted^b Odds Ratio (95% CI)	Pooled Unweighted^b Odds Ratio (95% CI)
Ln TUA1 ^c	1.87 (1.05, 3.36)	2.03 (1.22, 3.37)	2.29 (1.49, 3.51)	1.92 (1.00, 3.71)	1.87 (1.03, 3.44)	1.95 (1.32, 2.90)
Ln TUA2 ^d	1.26 (0.97, 1.66)	1.36 (0.99, 1.89)	1.67 (1.17, 2.40)	1.44 (0.89, 2.33)	1.40 (1.00, 1.97)	1.37 (1.04, 1.79)

^aModels adjusted for age, gender, race, family-poverty income ratio, BMI classification, and ln urinary creatinine. ^bModels adjusted for age, gender, race, family-poverty income ratio, BMI classification, ln urinary creatinine, and survey cycle. ^cTUA1 = As3 + As5 + MMA + DMA.

^dTUA2 = Total As – AsB – AsC.

Figure Legend

Figure 1. ^aAdjusted odds ratio from penalized splines models for negative VZV IgG by total urinary arsenic: A) TUA1 unweighted, B) TUA2 unweighted, C) TUA1 weighted and (D) TUA2 weighted. ^aOdds ratio based on penalized splines for ln transformed total arsenic exposure. Models fully adjusted for age, ln urinary creatinine, sex, race/ethnicity, family poverty-income ratio, BMI classification and survey cycle. A and B are unrestricted splines with 95% confidence intervals, C and D were estimated from weighted models restricted to knots at 5th, 25th, 50th, 95th percentiles of exposure.

Figure 1.

